

# Lecture 8: Genetic variants and their interpretation

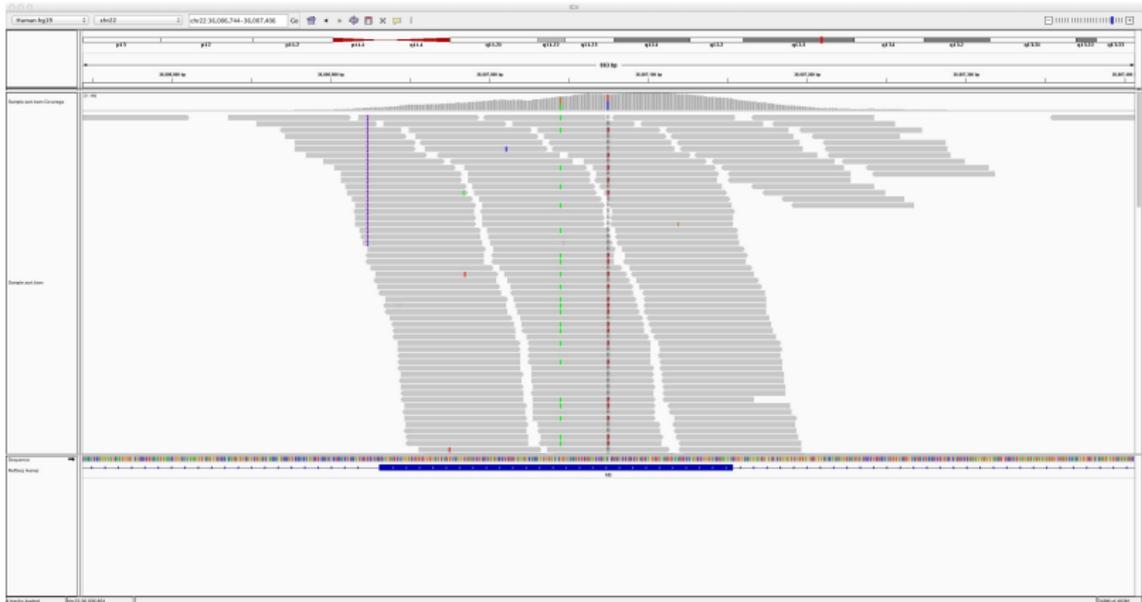
Paolo Provero

# NGS and genetic variants

NGS techniques allow us to detect the *genetic variants* that characterize each individual

- *Targeted sequencing*: detect variants in selected regions
  - Whole Exome Sequencing
  - Sequencing of candidate genes or exons
  - The smaller the target the greater the sequencing *depth* with the same number of reads
- *Whole genome sequencing*

# Variant detection



# VCF format

To save space, the variants are expressed as *differences* from the reference sequence

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##fileformat=VCFv4.0
##fileDate=20090805
##source=myImputationProgramV3.1
##reference=1000GenomesPilot-NCBI36
##phasing=partial
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##INFO=<ID=DP,Number=1,Type=Integer,Description="Total Depth">
##INFO=<ID=AF,Number=.,Type=Float,Description="Allele Frequency">
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##INFO=<ID=H2,Number=0,Type=Flag,Description="HapMap2 membership">
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##FORMAT=<ID=DP,Number=1,Type=Integer,Description="Read Depth">
##FORMAT=<ID=HQ,Number=2,Type=Integer,Description="Haplotype Quality">
#CHROM POS ID REF ALT QUAL FILTER INFO FORMAT NA00001 NA00002 NA00003
20 14370 rs6054257 G A 29 PASS NS=3;DP=14;AF=0.5;DB;H2 GT:GQ:DP:HQ 0|0:48:1:51,51 1|0:48:8:51,51 1/1:43:5:..
20 17330 . T A 3 q10 NS=3;DP=11;AF=0.017 GT:GQ:DP:HQ 0|0:49:3:58,50 0|1:3:5:65,3 0/0:41:3
20 1110696 rs6040355 A G,T 67 PASS NS=2;DP=10;AF=0.333,0.667;AA=T;DB GT:GQ:DP:HQ 1|2:21:6:23,27 2|1:2:0:18,2 2/2:35:4
20 1230237 . T . 47 PASS NS=3;DP=13;AA=T GT:GQ:DP:HQ 0|0:54:7:56,60 0|0:48:4:51,51 0/0:61:2
20 1234567 microsat1 GTCT G,GTACT 50 PASS NS=3;DP=9;AA=G GT:GQ:DP 0/1:35:4 0/2:17:2 1/1:40:3
```

# Variant interpretation

Problem: determine which variants are causal of a phenotype/disease

- *Mendelian* phenotypes
  - *rare* variants
  - high *penetrance*
  - in coding regions
- *Complex* phenotypes
  - *common* variants
  - non-coding regions

## Effect on the phenotype

The likelihood that a variant will affect the phenotype depends on where the variant is located (coding exon, UTR, intron, intergenic)

- Coding variants can be classified based on their effect on the protein sequence
  - synonymous: no change in protein sequence (possible due to the degenerate nature of the genetic code)
  - missense: changes one amino acid into another
    - conservative: new aa is chemically similar to old one
    - non-conservative: new aa is chemically different – likely change in structure
  - nonsense: introduces a stop codon
- Non-coding variants are assumed to affect phenotype by changing gene regulation
  - e.g. by creating/destroying a transcription factor binding site
  - much more difficult to classify/interpret

# Coding variants: the XLMR example

ARTICLES

nature  
genetics

## A systematic, large-scale resequencing screen of X-chromosome coding exons in mental retardation

Patrick S Tarpey<sup>1</sup>, Raffaella Smith<sup>1</sup>, Erin Pleasance<sup>1</sup>, Annabel Whibley<sup>2</sup>, Sarah Edkins<sup>1</sup>, Claire Hardy<sup>1</sup>, Sarah O'Meara<sup>1</sup>, Calli Latimer<sup>1</sup>, Ed Dicks<sup>1</sup>, Andrew Menzies<sup>1</sup>, Phil Stephens<sup>1</sup>, Matt Blow<sup>1</sup>, Chris Greenman<sup>1</sup>, Yali Xue<sup>1</sup>, Chris Tyler-Smith<sup>1</sup>, Deborah Thompson<sup>3</sup>, Kristian Gray<sup>1</sup>, Jenny Andrews<sup>1</sup>, Syd Bartherpe<sup>1</sup>, Gemma Buck<sup>1</sup>, Jennifer Cole<sup>1</sup>, Rebecca Dunmore<sup>1</sup>, David Jones<sup>1</sup>, Mark Maddison<sup>1</sup>, Tatiana Mironenko<sup>1</sup>, Rachel Turner<sup>1</sup>, Kelly Turrell<sup>1</sup>, Jennifer Varian<sup>1</sup>, Sofie West<sup>1</sup>, Sara Widaa<sup>1</sup>, Paul Wray<sup>1</sup>, Jon Teague<sup>1</sup>, Adam Butler<sup>1</sup>, Andrew Jenkinson<sup>1</sup>, Mingming Jia<sup>1</sup>, David Richardson<sup>1</sup>, Rebecca Shepherd<sup>1</sup>, Richard Wooster<sup>1</sup>, M Isabel Tejada<sup>4</sup>, Francisco Martinez<sup>5</sup>, Gemma Carvill<sup>6</sup>, Rene Goliath<sup>6</sup>, Arjan P M de Brouwer<sup>7</sup>, Hans van Bokhoven<sup>7</sup>, Hilde Van Esch<sup>8</sup>, Jamel Chelly<sup>9</sup>, Martine Raynaud<sup>10</sup>, Hans-Hilger Ropers<sup>11</sup>, Fatima E Abidi<sup>12</sup>, Anand K Srivastava<sup>12</sup>, James Cox<sup>2</sup>, Ying Luo<sup>2</sup>, Uma Mallya<sup>2</sup>, Jenny Moon<sup>2</sup>, Josef Parnau<sup>2</sup>, Shehla Mohammed<sup>13</sup>, John L Tolmie<sup>14</sup>, Cheryl Shoubridge<sup>15</sup>, Mark Corbett<sup>15</sup>, Alison Gardner<sup>15</sup>, Eric Haan<sup>15</sup>, Simitdhorn Rujirabanjerd<sup>15</sup>, Marie Shaw<sup>15</sup>, Lucianne Vandeleur<sup>15</sup>, Tod Fullston<sup>15</sup>, Douglas F Easton<sup>3</sup>, Jackie Boyle<sup>16</sup>, Michael Partington<sup>16</sup>, Anna Hackett<sup>16</sup>, Michael Field<sup>16</sup>, Cindy Skinner<sup>12</sup>, Roger E Stevenson<sup>12</sup>, Martin Bobrow<sup>2</sup>, Gillian Turner<sup>16</sup>, Charles E Schwartz<sup>12</sup>, Jozef Gecz<sup>15,17</sup>, F Lucy Raymond<sup>2</sup>, P Andrew Futreal<sup>1</sup> & Michael R Stratton<sup>1,18</sup>

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*Nature Genetics* 2009

## Coding variants: the XLMR example

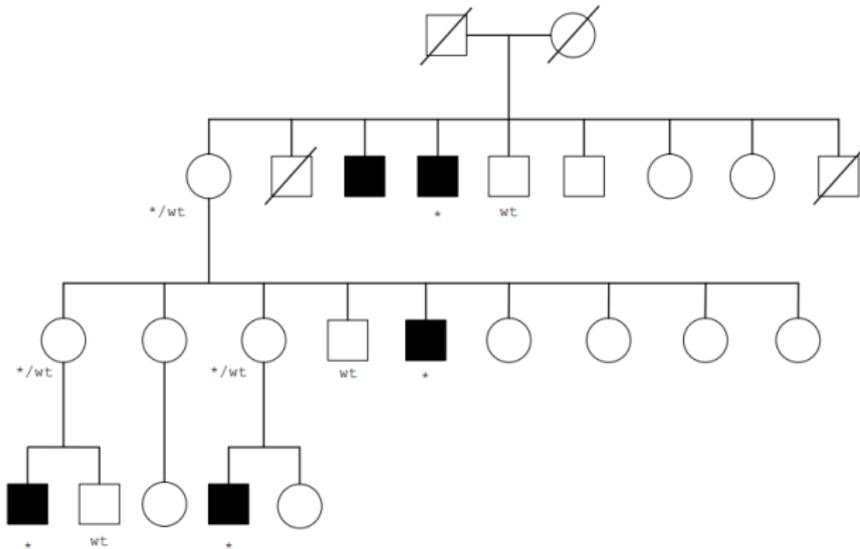
- 208 families with MR and pattern of transmission compatible with X linkage
- Exome sequencing of ~700 genes in X chromosome
- Most differences in coding sequence between individuals are recurring and found in dbSNP
- Consider *truncating* variants only (found in 30 genes)

# Segregation

A variant can be *causal* of the disease only if it *segregates* - presence of the variant in all affected members and only in affected members

## Family 62

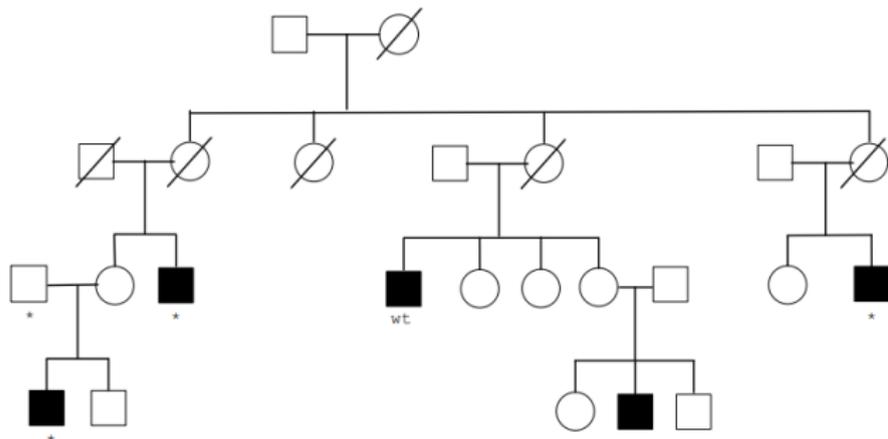
UPF3B(NM\_080632.1) c.1288C>T p. R430X



# Non-segregation

## Family 74

ITIH5L (NM\_198510.1) IVS7+1ins T



# Message

- Exome sequencing can help finding causal variants of Mendelian diseases
- However even truncating variants can be compatible with normal phenotype
  - “loss of function of 1% of the genes in the X chromosome is compatible with apparently normal existence”
- Another recent study found an average of ~40 homozygous LOF mutations in normal individuals
- Therefore when sequencing the exome of a *proband* with a genetic disease we expect to find many candidate mutations
  - Need for *variant prioritization*

# Variant prioritization

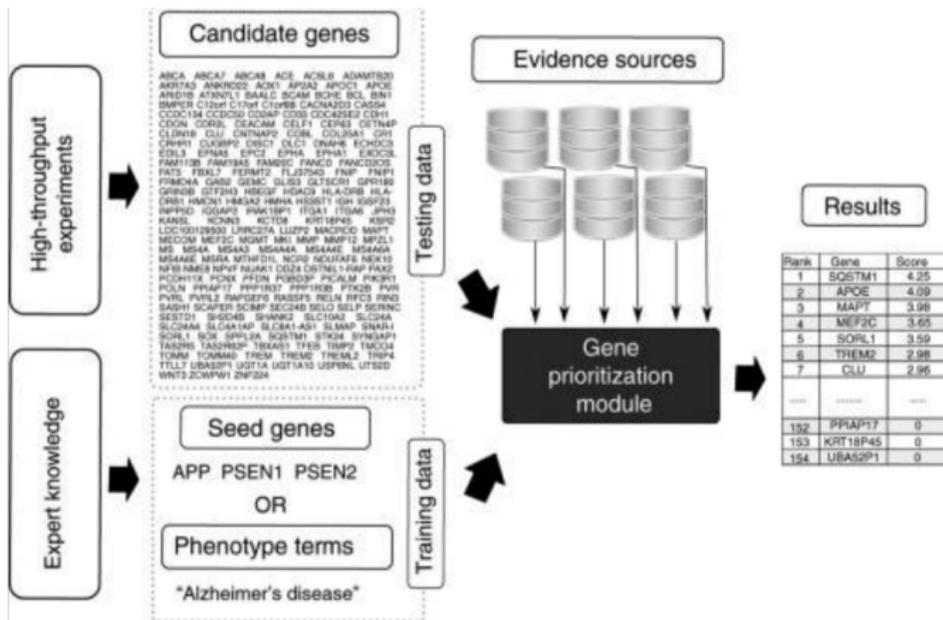
Problem:

- Among all the variants found in a proband with a genetic disease, find the ones most likely to be causative

The following criteria increase the probability that a variant is causative

- very low frequency in the population
- predicted strong effect on the protein
- gene involved in biological process relevant to the disease
- gene interacts with genes involved in the same/similar diseases

# Variant prioritization



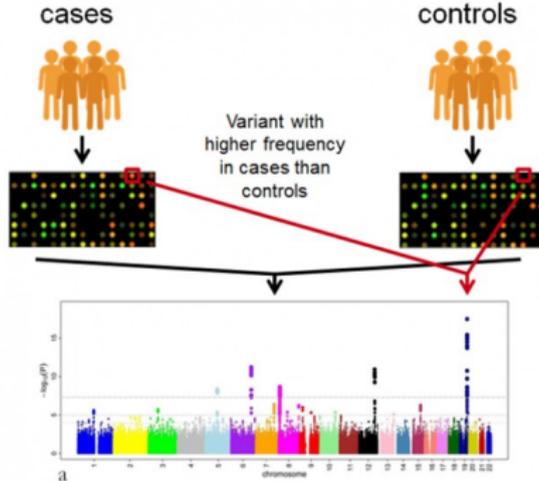
From Zolotareva & Kleine, *J Integr Bioinform* 16:20180069 (2019).

# Complex phenotypes

- Complex trait: determined by both genetic and non-genetic (e.g. environmental, behavioral, . . . ) factors
  - Diabetes
  - Rheumatoid arthritis
  - Crohn disease
  - Height
  - Blood pressure
  - Lymphocyte count
- Genetic determinants are usually
  - Many variants of small effect
  - Non-coding (hence presumably regulatory)

# GWAS

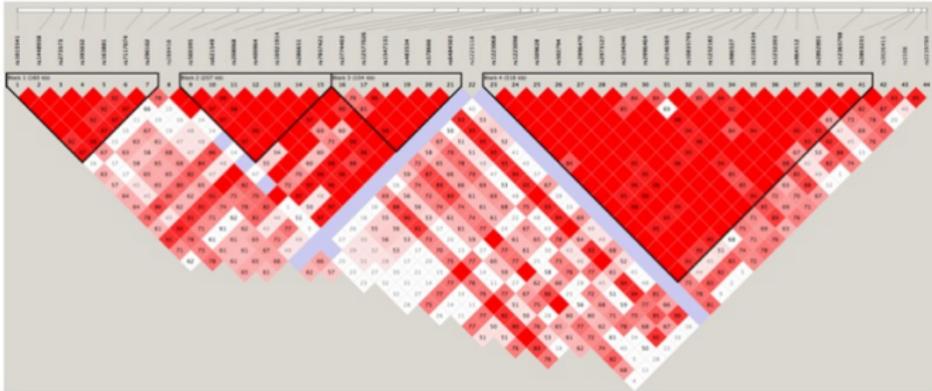
Genome-wide associations studies look for variants with significantly different frequency in cases and controls



# Predicting vs understanding complex traits

- GWAS hits can be used to build *polygenic scores* predicting disease risk
- To understand disease (hence possibly find cures) one needs to understand the mechanism leading from variant to disease
- GWAS hits are mostly non-coding → regulatory effect
  - What is the *target* of a regulatory variant (gene whose expression is altered by the variant)?
  - What is the effect of the variant?
    - Regulatory code much less understood than genetic code
  - Which one is the causal variant?
    - *Linkage disequilibrium* implies that many neighboring variants are inherited together, thus showing the same correlation with the disease although presumably only one or a few are causative

# Linkage disequilibrium



Strong correlation between nearby SNPs indicate that they are inherited together (no recombination events)



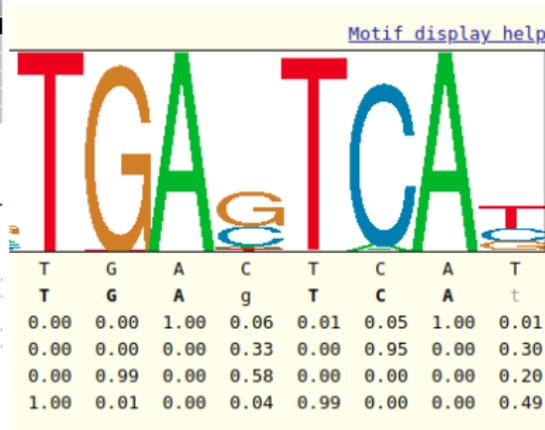
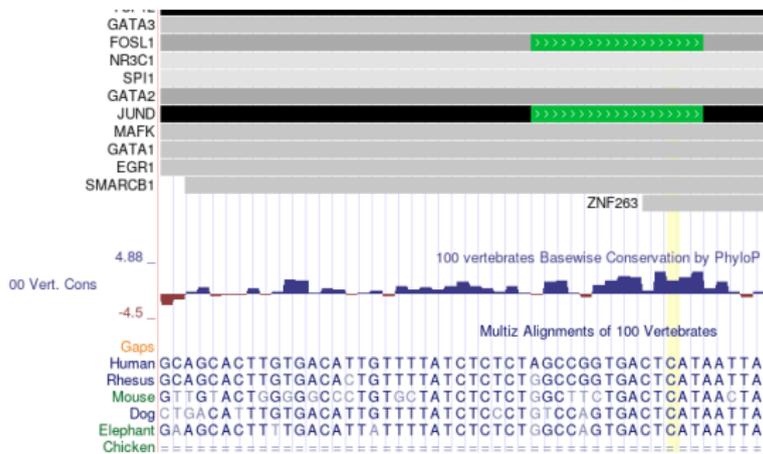
# An example: rs17293632

- Colocalizes with several transcription factor binding peaks from ChIP-seq
- In particular, is within the core binding motif of AP-1



# An example: rs17293632

The variant disrupts a highly conserved AP-1 motif



# Interpreting rs17293632

- The variant rs17293632 disrupts a highly conserved (hence, probably functional) AP-1 binding site
- Thus altering gene expression, presumably of SMAD3
- Thus altering the phenotype, e.g. by conferring susceptibility to Crohn's disease

## Genetic and epigenetic fine mapping of causal autoimmune disease variants

Kyle Kai-How Farh<sup>1,2\*</sup>, Alexander Marson<sup>3\*</sup>, Jiang Zhu<sup>1,4,5,6</sup>, Markus Kleinewietfeld<sup>1,7†</sup>, William J. Housley<sup>7</sup>, Samantha Beik<sup>1</sup>, Noam Shores<sup>1</sup>, Holly Whitton<sup>1</sup>, Russell J. H. Ryan<sup>1,5</sup>, Alexander A. Shishkin<sup>1,8</sup>, Meital Hatan<sup>1</sup>, Marlene J. Carrasco-Alfonso<sup>9</sup>, Dita Mayer<sup>9</sup>, C. John Luckey<sup>9</sup>, Nikolaos A. Patsopoulos<sup>1,10,11</sup>, Philip L. De Jager<sup>1,10,11</sup>, Vijay K. Kuchroo<sup>12</sup>, Charles B. Epstein<sup>1</sup>, Mark J. Daly<sup>1,2</sup>, David A. Hafler<sup>1,7§</sup> & Bradley E. Bernstein<sup>1,4,5,6§</sup>

# eQTLs

To understand GWAS hits we need a systematic analysis of the effect of variants on gene expression.

An eQTL (expression quantitative trait locus) is a variant significantly associated with the expression of a gene

- gene expression considered as a *quantitative trait*, like height or lymphocyte count

An eQTL study needs a large cohort (hundreds of individuals) for which we have

- expression data
- genotyping (i.e. *dosage* [0,1, or 2] of each genetic variant)

The analysis is performed by *linear regression*

# Regression

- *Regression* is the main statistical method for the study of the *dependence* among two (or more) variables  $x$  and  $y$
- Assumes the existence of a *regression function*  $f(x)$  which represents the true dependence of  $y$  on  $x$  as

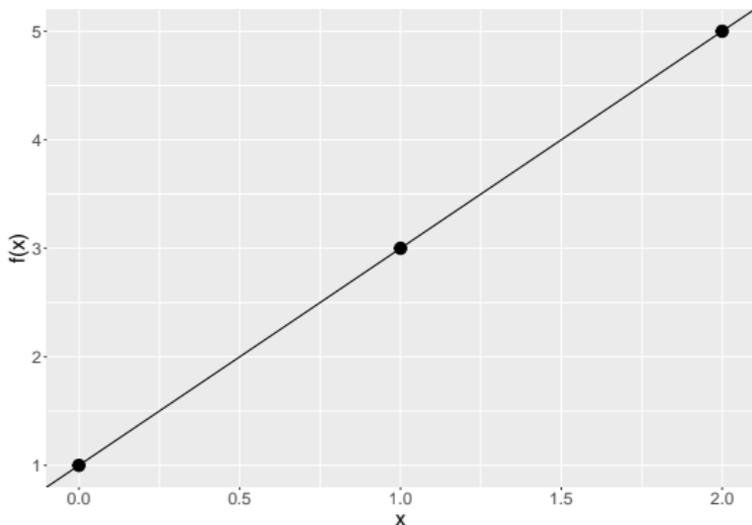
$$y = f(x) + \epsilon$$

- In our case:
  - $x$ : dosage of a variant ( $x \in \{0, 1, 2\}$ )
  - $y$ : expression of a gene
- The *error term*  $\epsilon$  represents random fluctuations, or the effects of variables that we do not consider
  - e.g. environmental effects on the expression  $y$  of the gene

# Linear regression

In *linear regression* we assume  $f(x)$  to be a straight line:

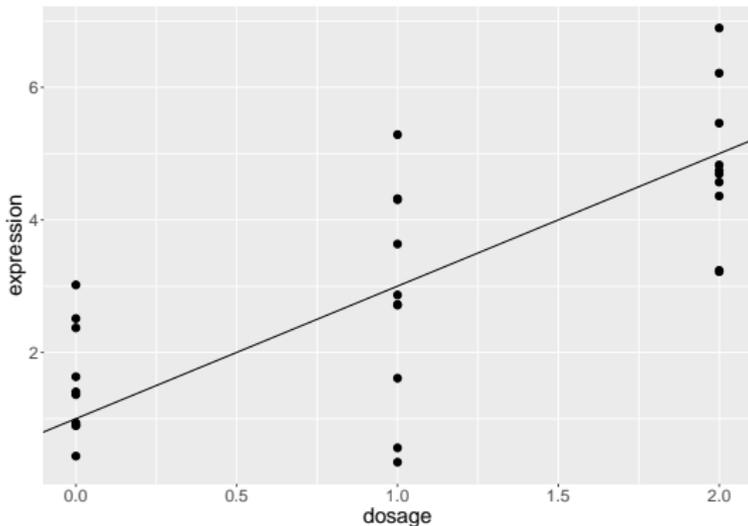
$$f(x) = \beta_0 + \beta_x x$$



In our case this means that we consider the effects of the paternal and maternal alleles on gene expression as independent and additive.

# Error term

If the regression function is linear we expect, considering the error term  $\epsilon$ , the actual measurements to look like this



## Estimating $\beta_0$ and $\beta_x$

Given the data:

- dosage  $x_i$  ( $i = 1 \dots N$ ) and expression  $y_i$  ( $i = 1 \dots N$ ) for a large number  $N$  of individuals

our first goal is to estimate the regression function, i.e. the values of  $\beta_0$  and  $\beta_x$ .

This is done by choosing the values that minimize the *mean square error*

$$MSE = \sum_{i=1}^N (y_i - f(x_i))^2 = \sum_{i=1}^N (y_i - \beta_0 - \beta_x x_i)^2$$

## Is this an eQTL?

Regression function:

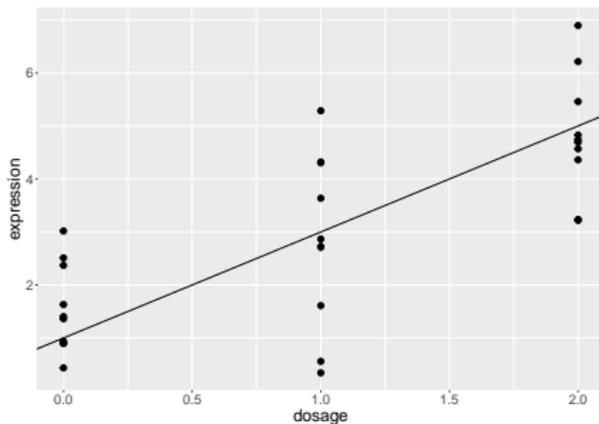
$$f(x) = \beta_0 + \beta_x x$$

The variant is called an eQTL for the gene if  $\beta_x \neq 0$  (if  $\beta_x = 0$  expression does *not* actually depend on the variant dosage).

- Null hypothesis:  $\beta_x = 0$
- Regression algorithms provide us not only with the value of  $\beta_x$  (and  $\beta_0$ ) but also with their *uncertainties*
- These can be used to *test* the null hypothesis and obtain a *P*-value
- Small *P*-values indicate that
  - the null hypothesis is probably false
  - that is,  $\beta_x \neq 0$
  - that is, the variant dosage does indeed affect the expression of the gene
  - that is, the variant is an eQTL of the gene

# Example

For our (fake) data



- the estimated  $\beta_x$  is 1.64
- with an uncertainty (standard error) of 0.277
- so that it is very unlikely that the true  $\beta_x$  is 0
- indeed the  $P$ -value is  $2.36 \cdot 10^{-6}$ .

# The GTEx project

## The GTEx Consortium atlas of genetic regulatory effects across human tissues

[THE GTEx CONSORTIUM](#)

*SCIENCE* • 11 Sep 2020 • Vol 369, Issue 6509 • pp. 1318-1330 • DOI:10.1126/science.aaz1776

- eQTLs can be tissue-specific
- e.g. by altering the binding site of a brain-specific TF
- GTEx: eQTL analysis in ~50 human tissues
  - analyze all variants within 1 Mb of each gene

# Back to rs17293632

GTEx confirms that rs17293632 is an eQTL of SMAD3 in two tissues (but also of other neighboring genes)

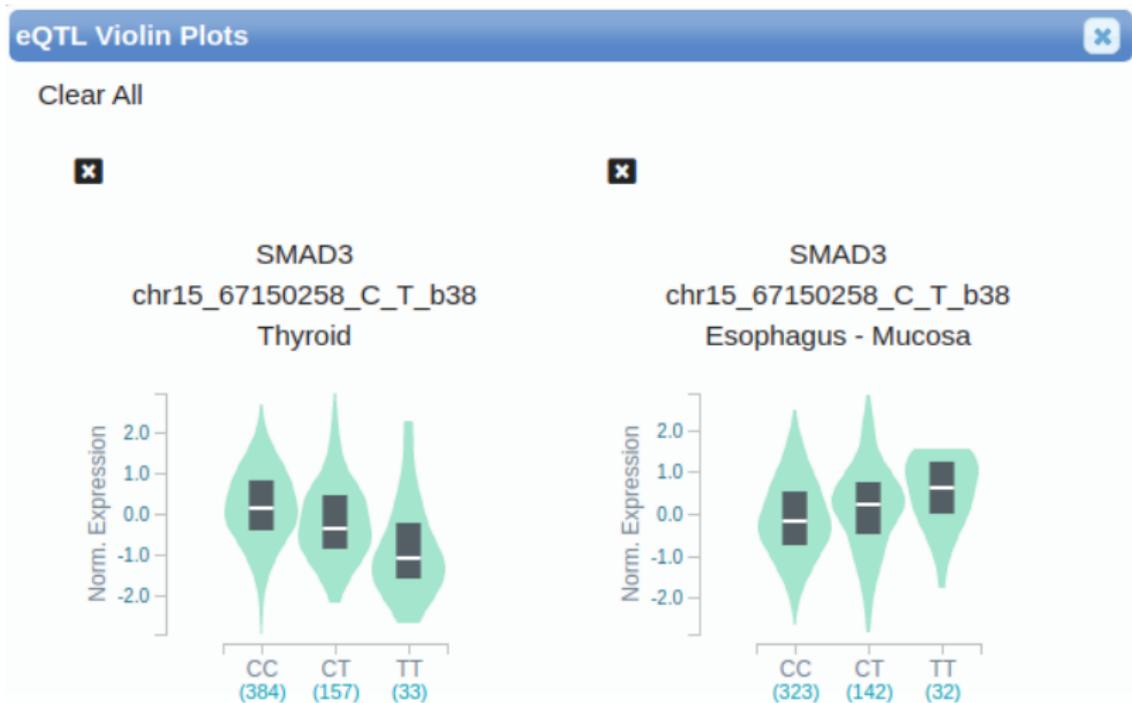
eQTLs of chr15\_67150258\_C\_T\_b38

Copy	CSV					
Gencode Id	Gene Symbol	Variant Id	SNP	P-Value	NES	Tissue
ENSG00000166949.15	SMAD3	chr15_67150258_C_T_b38	rs17293632 dbSNP	5.8e-30	-0.48	Thyroid
ENSG00000166949.15	SMAD3	chr15_67150258_C_T_b38	rs17293632 dbSNP	0.0000038	0.22	Esophagus - Mucosa
ENSG00000103591.12	AAGAB	chr15_67150258_C_T_b38	rs17293632 dbSNP	0.000017	-0.13	Whole Blood
ENSG00000103591.12	AAGAB	chr15_67150258_C_T_b38	rs17293632 dbSNP	0.000064	-0.10	Esophagus - Mucosa
ENSG00000033800.13	PIAS1	chr15_67150258_C_T_b38	rs17293632 dbSNP	0.00024	0.094	Thyroid

Showing 1 to 5 of 5 entries

# Tissue-dependent effects

The effect of the variant on SMAD3 expression depends on the tissue:



# GWAS and eQTLs

- eQTLs are useful in interpreting variants found by GWAS to be associated to complex traits/diseases
- Enrichment of eQTLs has been found among GWAS hits
- Recently a more global approach to the use of eQTLs has emerged: intermediate molecular phenotypes
  - In particular, *transcriptome-wide association studies* (TWAS)

# Gene expression as an intermediate molecular phenotypes

GWAS:

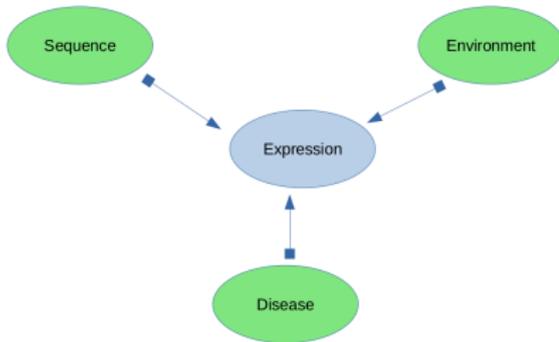


As most hits are non-coding, we actually believe



Gene expression is an *intermediate molecular phenotype*

# Components of gene expression



- Gene expression is not determined solely by genetics (sequence)
- However only the *genetic component of gene expression* (GREx) can mediate between variants and disease



# Transcriptome-wide association studies

nature  
genetics

## TECHNICAL REPORTS

A gene-based association method for mapping traits using reference transcriptome data

Eric R Gamazon<sup>1,2,5</sup>, Heather E Wheeler<sup>3,5</sup>, Kanaan P Shah<sup>1,5</sup>, Sahar V Mozaffari<sup>4</sup>, Keston Aquino-Michaels<sup>1</sup>, Robert J Carroll<sup>1</sup>, Anne E Eyster<sup>6</sup>, Joshua C Denny<sup>5</sup>, GTEx Consortium<sup>7</sup>, Dan L Nicolae<sup>1,4,5</sup>, Nancy J Cox<sup>1,2,4</sup> & Hae Kyung Im<sup>1</sup>

- Use eQTL data *from control individuals* (e.g. GTEx) to build a model predicting expression from genotype
- Use the models to compute the *GREx* of all genes for the GWAS diseased and control individuals
- Look for genes whose GREx correlates with the disease
- *Genetic variants affect the disease through the expression of these genes*
- The genes might be *therapeutic targets*

# Other molecular QTLs

Article | [Open Access](#) | Published: 19 September 2019

## Genome-wide identification of DNA methylation QTLs in whole blood highlights pathways for cardiovascular disease

Tianxiao Huan , Roby Joehanes, Ci Song, Fen Peng, Yichen Guo, Michael Mendelson, Chen Yao, Chunyu Liu, Jiantao Ma, Melissa Richard, Golareh Agha, Weihua Guan, Lynn M. Almli, Karen N. Conneely, Joshua Keefe, Shih-Jen Hwang, Andrew D. Johnson, Myriam Fornage, Liming Liang  & Daniel Levy 

*Nature Communications* 10, Article number: 4267 (2019) | [Cite this article](#)

5934 Accesses | 17 Citations | 3 Altmetric | [Metrics](#)

## Pooled ChIP-Seq Links Variation in Transcription Factor Binding to Complex Disease Risk

Ashley K. Tehranchi,<sup>1</sup> Marsha Myrthil,<sup>2</sup> Trevor Martin,<sup>1</sup> Brian L. Hie,<sup>3</sup> David Golan,<sup>4,5</sup> and Hunter B. Fraser<sup>1\*</sup>

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<http://dx.doi.org/10.1016/j.cell.2016.03.041>

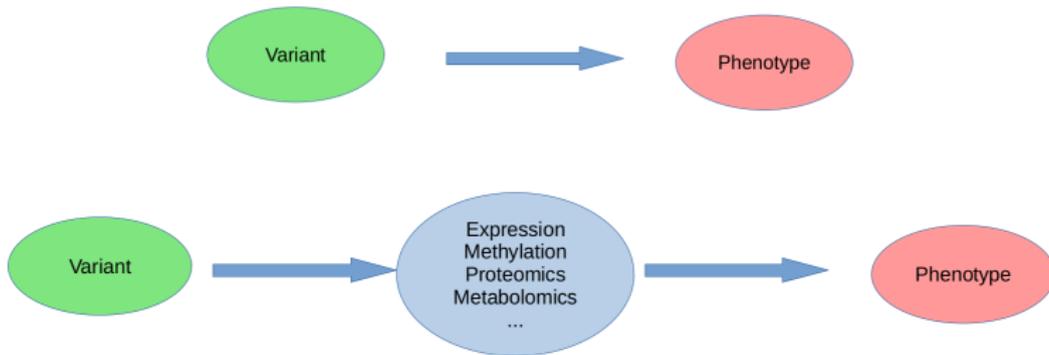
## A Genome-Wide Metabolic QTL Analysis in Europeans Implicates Two Loci Shaped by Recent Positive Selection

George Nicholson , Mattias Rantalainen, Jia V. Li, Anthony D. Maher, Daniel Malmodin, Kourosh R. Ahmadi, Johan H. Faber,

Amy Barrett, Josine L. Min, N. William Rayner, Henrik Toft, Maria Krestyaninova, Juris Viksna, [...], Chris C. Holmes 

[ [view all](#) ]

# Intermediate molecular phenotypes



# TWAS generalizations

Use other molecular phenotypes as intermediate phenotypes. . .

**PWAS: proteome-wide association study—linking genes and phenotypes by functional variation in proteins**

Nadav Brandes<sup>1\*</sup> , Nathan Linal<sup>1</sup> and Michal Linal<sup>2\*</sup>

. . . or even macroscopic phenotypes

**Imaging-wide association study: Integrating imaging endophenotypes in GWAS**

Zhiyuan Xu, Chong Wu, Wei Pan<sup>\*</sup>, for the Alzheimer's Disease Neuroimaging Initiative<sup>1</sup>

*Division of Biostatistics, School of Public Health, University of Minnesota, Minneapolis, MN 55455, USA*

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# Summary

Understanding the relationship between variants and phenotypes/diseases presents different challenges for Mendelian and complex phenotypes

- Mendelian diseases: find the causal coding variant through *variant prioritization*
- Complex diseases: understand the effect of many variants on gene regulation

In both cases the understanding of the causal relationship can in principle lead to new therapeutic strategies